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REVIEW

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Role of Mutations in NOD2/CARD15, ATG16L1, and IRGM in the Pathogenesis of Crohn's Disease

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Abstract

This review article summarizes the issues concerning the pathogenesis of Crohn's disease (CD) based on the results of large-scale genome-wide association studies. The role of defects in innate immunity associated with mutations in specific genes such as those regulating bacterial pattern recognition (*NOD2/CARD15*) and autophagy/xenophagy (*ATG16L1* and *IRGM*) in CD is also discussed. Basic pathogenetic hypotheses that aim to interpret the association between specific gene mutations and CD development are presented.

Keywords: Crohn's disease, innate immunity, autophagy, xenophagy, microbiota, NOD2/CARD15, ATG16L1, IRGM

Introduction

Crohn's disease (CD) is a multisystemic disease with unknown etiology and is characterized by nonspecific granulomatous transmural inflammation with segmented injuries in any part of the gastrointestinal tract (GIT) [1, 2]. Epidemiological data shows that the incidence and prevalence of CD in northern Europe and North America have increased, which also has major social impact [3, 4]. In last decades, the significant increase in understanding of molecular genetics of CD, allowed the identification of genetic determinants associated with CD pathology [2, 5].

The results of various epidemiological studies have identified the role of genetic factors in the pathogenesis of CD. In particular, several observations provide evidence that this specific pathology commonly occurs in families of patients with CD [1, 2, 6, 7]. Approximately one of every five patients with CD has at least one relative with the same pathology [8]. However, in studies involving monozygotic twins, a 67% concordance (comorbidity in both twins) has been demonstrated for CD [2]. In addition, the ethnic variations in CD incidence rate are in favor of a genetic component in its pathogenesis. For example, the CD incidence rate is 2–4 times higher in Eastern Europe, with the exception of the Ashkenazi Jews and other ethnic groups [9]. Therefore, the enumerated factors serve as prerequisites for the identification of CD susceptible loci.

The evolution of molecular genetic technologies has resulted in a new epoch of genome-wide association studies (GWAS), which facilitates in the identification of the new candidate genes for CD based on their genomic localization, without reference to its biological effect or mode of inheritance [10]. GWAS have become the all-powerful catalyst in understanding the pathogenesis of CD, resulting in the identification of approximately more than 30 loci that confer a predisposition to CD [11, 12]. The most associations between CD and specific mutations include those regulating the bacterial pattern recognition and autophagy [2, 6, 13].

NOD2/CARD15. A first susceptibility locus identified for CD is *NOD2*, also known as *CARD15*; this gene was identified by two independent research groups in early 2001 [14,15]. This particular investigation could be considered as the most significant achievement in genetic studies of inflammatory bowel diseases [1,5].

NOD2/CARD15 encodes for the cytosolic protein, NOD2, which is an intracellular pattern-recognizing receptor binding to muramyl dipeptide (MDP) [16,17]. The last one is the component of bacterial peptidoglycan from walls of Gram-positive and Gram-negative bacteria [17]. NOD2 is preferentially expressed in immunocompetent cells such as macrophages, dendritic cells, and Paneth cells [18]. Paneth

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cells are specialized cells of the small intestines that produce antibacterial substances called α -defensins [19].

NOD2 consists of two CARD domains, one nucleotide binding domain (NBD), and a leucine-rich repeat region (LRR), which serves as the domain for MDP recognition [20]. Pattern recognition by NOD2 is associated with activation of signal transduction, which results in the induction of the transcription factor, NF- κ B. Its activation results in the transcription of several pro-inflammatory genes [17, 21, 22]. NOD2 is the most important component of innate immunity, which subsequently confirms the hypothesis that an alteration in the immune response to gastrointestinal microbiota influences the development of CD [2].

At least 30 alleles of *NOD2/CARD15* have been identified [23]. However, the most frequent allele variants in European and American populations that are associated with CD include two missense mutations, Arg702Trp, Gly908Arg, and a frameshift mutation Leu1007fsinsC [2, 6, 13-15]. These particular mutations represent 82% of all *NOD2/CARD15* gene variations that have been associated with CD [24].

The specific mutations in *NOD2/CARD15* affect the LRR–MDP recognition region and disrupt the binding of MDP to the NOD2 protein [25]. However, no unifying model currently explains the association between *NOD2/CARD15* mutations and CD pathogenesis.

Following the one of hypothesis the proinflammatory mediators' transcription and inflammation induction even under minimal bacterial invasion is explained by that the *NOD2/CARD15* gene mutations leads to hyperamplification in response to MDP stimulation in signal transduction through NF-κB [26]. On the over side that is out of this model frames it has been demonstrated for today that as minimum the *NOD2/CARD15* gene missense mutations functionally inactivate the protein [27].

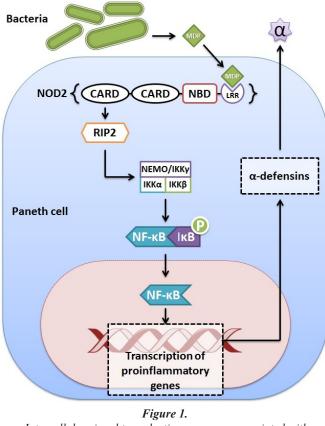
Another theory on CD pathogenesis is that NOD2 inhibits TLR2-associated signal pathways, thereby inducing the synthesis of proinflammatory cytokines such as IL-12 [28]. Mutations in *NOD2/CARD15* result in the upregulation of IL-12, followed by Th1-type adaptive immunity polarization [29].

Another hypothesis is that *NOD2/CARD15* mutations lead to not only changes in MDP recognition but also to a downregulation of α -defensin production by Paneth cells (Fig. 1) [18]. However, patients with CD and *NOD2/CARD15* mutations show a 50% decrease in α -defensin HD5 expression [30, 31]. These changes disrupt the homeostasis between the macro-organism and bowel microbiota [32]. The decrease in the protective potential results in quantitative and qualitative alterations in the microbiotic content, with an expansion of pathogenic and commensal bacteria, which mediates their translocation into the mucosal layer followed by the activation of adaptive immunity that primarily involves antigenpresenting cells (APC) and T-lymphocytes [32, 33].

Current clinical trials have demonstrated that *NOD2/ CARD15* gene polymorphisms are associated with early CD onset with inflammation, particularly involving the ileum (ileitis), as well as the early formation of strictures [34-36].

A previous report has shown that 20%–30% of patients

with CD harbor *NOD2/CARD15* mutations [2]. However, the penetrance of these variants is less than 1%; thus, these variations can occur even in individuals without CD [37]. Therefore, *NOD2/CARD15* mutations are not exclusively responsible for CD development.



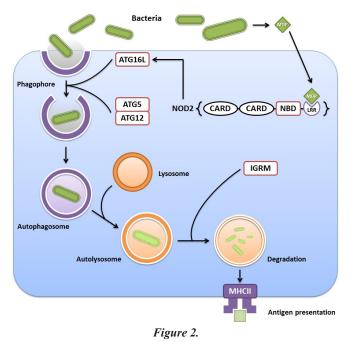
Intracellular signal transduction processes associated with α-defensin production in Paneth cells

Autophagy genes (ATG16L1 and IGRM)

Subsequent investigations in CD genetics have implicated autophagy elements [2, 6, 13, 38]. Autophagy is a cell mechanism involving excessive or damaged proteins, protein complexes, and performing cell organelles utilization by cell own lysosomes [39, 40]. Autophagy plays an important role in immune protection against viral, bacterial, and parasitic infections by selective microorganisms' utilization in lysosomes, called xenophagy [41]. Autophagy also serves as an integral link between innate and adaptive immunity, providing antigens with major histocompatibility complexes class II for the presentation (MHC-II) [42].

According to results of some GWAS, single nucleotide polymorphisms in *ATG16L1* (T300A) are associated with high-risk CD development [11, 43, 44]. The *ATG16L1* product is an autophagy modulating protein, ATG16L1, which forms a complex with ATG5-ATG12 and is responsible for autophagosome formation (Fig. 2) [45]. The role of NOD2 in autophagy induction in association with ATG16L1 has been established, although there is still a need to establish the significance between *NOD2/CARD15* gene mutations and CD pathogenesis [46, 47].

In general, ATG16L1 mutations are believed to be associated with alterations in xenophagy activation based on the recognition of intracellular bacteria patterns by NOD2 [48]. In addition to the ATG16L1 defects that lead to bowel deregulation, Paneth α -defensin exocytosis may also play a direct role in CD development [49].



The main stages of xenophagy (autophagy)

GWAS have identified another autophagy gene, *IRGM*, which has been associated with CD [11, 50]. This gene encodes the immunity-related GTPase family M protein (IRGM) [51]. The actual function and role of this protein is unknown. It has been suggested that IRGM plays a role in protecting the cell against intracellular bacteria by inducing autolysosome substrate degradation [52]. Defects in autophagy play a major role in CD pathogenesis. These defects result in an insufficiency in the clearance of pathogens in the GIT and interruptions in the activation of adaptive immunity involving MHC-II molecule antigen presentation [38].

Conclusion

The last few decades have been marked by major breakthroughs in understanding the molecular genetic basis of CD. Results of large-scale GWAS have substantially changed our view on the pathogenesis of CD. Alterations in innate immunity, which influence autophagy and the recognition of microbiota structures, appear to be the most important stage of CD pathogenesis, followed by the irrational activation of adaptive immunity components.

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Competing interests

The authors declare that they have no competing interests.

References

1. Vorob'ev GI, Khalif IL. Unspecific inflammatory bowel diseases. Moscow: Miklosh, 2008.

2. Sands BE, Siegel CA. Crohn's disease. In: Feldman M, Friedman LS, Brandt LJ, eds. Sleisenger & Fordtran's Gastrointestinal and Liver Disease. 9th ed. Philadelphia, Pa: Saunders Elsevier; 2010: chap 111.

3. Burisch J, Munkholm P. Inflammatory bowel disease epidemiology. Curr Opin Gastroenterol 2013; 29 (4): 357-62.

4. Kappelman MD, Rifas-Shiman SL, Kleinman K, Ollendorf D, Bousvaros A, Grand RJ, et al. The prevalence and geographic distribution of Crohn's disease and ulcerative colitis in the United States. Clin Gastroenterol Hepatol 2007; 5:1424-9.

5. Geremia A, Biancheri P, Allan P, Corazza GR, Di Sabatino A. Innate and adaptive immunity in inflammatory bowel disease. Autoimmun Rev 2014; 13(1):3-10.

6. Baumgart DC, Sandborn WJ. Crohn's disease. Lancet 2012; 380 (9853): 1590-1605.

7. Peeters M, Nevens H, Baert F, Hiele M, de Meyer AM, Vlietinck R, et al. Familial aggregation in Crohn's disease: Increased age-adjusted risk and concordance in clinical characteristics. Gastroenterology 1996; 111:597-603.

8. Dorn SD, Abad JF, Panagopoulos G, Korelitz BI. Clinical characteristics of familial versus sporadic Crohn's disease using the Vienna Classification. Inflamm Bowel Dis 2004; 10: 201-206.

9. Yan B, Panaccione R, Sutherland L. I am Jewish: what is my risk of developing Crohn's disease? Inflamm Bowel Dis 2008; 14 Suppl. 2:S26-7.

10. Xavier RJ, Rioux JD. Genome-wide association studies: A new window into immune-mediated diseases. Nature Rev Immunol 2008; 8:631-43.

11. Barrett JC, Hansoul S, Nicolae DL, Cho JH, Duerr RH, Rioux JD, et al. Genome-wide association defines more than 30 distinct susceptibility loci for Crohn's disease. Nat Genet 2008; 40:955-62.

12. Lee YH, Song GG. Pathway analysis of a genome-wide association study of ileal Crohn's disease. DNA Cell Biol 2012; 31(10):1549-54.

13. Tsianos EV, Katsanos KH, Tsianos VE. Role of genetics in the diagnosis and prognosis of Crohn's disease. World J Gastroenterol 2012; 18(2):105-18.

14. Hugot JP, Chamaillard M, Zouali H, Lesage S, Cézard JP, Belaiche J, et al. Association of NOD2 leucine-rich repeat variants with susceptibility to Crohn's disease. Nature 2001; 411:599-603.

15. Ogura Y, Bonen DK, Inohara N, Nicolae DL, Chen FF, Ramos R, et al. A frameshift mutation in NOD2 associated with susceptibility to Crohn's disease. Nature 2001; 411:603-6. 16. Girardin SE, Boneca IG, Viala J, Chamaillard M, Labigne A, Thomas G, et al. Nod2 is a general sensor of peptidoglycan through muramyl dipeptide (MDP) detection. J Biol Chem 2003; 278(11):8869-72.

17. Grimes CL, Ariyananda Lde Z, Melnyk JE, O'Shea EK. The innate immune protein Nod2 binds directly to MDP, a bacterial cell wall fragment. J Am Chem Soc 2012; 134(33):13535-7.

18. Lala S, Ogura Y, Osborne C, Hor SY, Bromfield A, Davies S, et al. Crohn's disease and the NOD2 gene: A role for paneth cells. Gastroenterology 2003; 125:47-57.

19. Ayabe T, Satchell DP, Wilson CL, Parks WC, Selsted ME,

Ouellette AJ. Secretion of microbicidal alpha-defensins by intestinal Paneth cells in response to bacteria. Nat Immunol 2000; 1:113–118

20. Ogura Y, Inohara N, Benito A, Chen FF, Yamaoka S, Nunez G. Nod2, a Nod1/Apaf-1 family member that is restricted to monocytes and activates NF-kappaB. J Biol Chem 2001; 276(7):4812-8.

21. Tattoli I, Travassos LH, Carneiro LA, Magalhaes JG, Girardin SE. The Nodosome: Nod1 and Nod2 control bacterial infections and inflammation. Semin Immunopathol 2007; 29(3):289-301.

22. Lécine P, Esmiol S, Métais JY, Nicoletti C, Nourry C, McDonald C, et al. The NOD2-RICK complex signals from the plasma membrane. J Biol Chem 2007; 282(20):15197-207. 23. Lesage S, Zouali H, Cézard JP, Colombel JF, Belaiche J, Almer S, et al. CARD15/NOD2 mutational analysis and genotype-phenotype correlation in 612 patients with inflammatory bowel disease. Am J Hum Genet 2002; 70:845–57. 24. Yazdanyar S, Weischer M, Nordestgaard BG. Genotyping for NOD2 genetic variants and crohn disease: a metaanalysis. Clin Chem 2009; 55(11):1950-7.

25. Yamamoto S, Ma X. Role of Nod2 in the development of Crohn's disease. Microbes Infect 2009; 11(12): 912-918.

26. Maeda S, Hsu LC, Liu H, Bankston LA, Iimura M, Kagnoff MF, et al. Nod2 mutation in Crohn's disease potentiates NF-kappaB activity and IL-1beta processing. Science 2005; 307(5710):734-8.

27. Vermeire S. Review article: genetic susceptibility and application of genetic testing in clinical management of inflammatory bowel disease. Aliment Pharmacol Ther 2006; 24 Suppl 3:2-10.

28. Yang Z, Fuss IJ, Watanabe T, Asano N, Davey MP, Rosenbaum JT, et al. NOD2 transgenic mice exhibit enhanced MDP-mediated down-regulation of TLR2 responses and resistance to colitis induction. Gastroenterology 2007; 133: 1510–1521.

29. Watanabe T, Kitani A, Murray PJ, Strober W. NOD2 is a negative regulator of Toll-like receptor 2-mediated T helper type 1 responses. Nat Immunol 2004; 5(8):800-8.

30. Wehkamp J, Salzman NH, Porter E, Nuding S, Weichenthal M, Petras RE, et al. Reduced paneth cell α -defensins in ileal Crohn's disease. Proc. Natl Acad Sci USA 2005; 102(50), 18129-34. 31. Wehkamp J, Harder J, Weichenthal M, Schwab M, Schäffeler E, Schlee M, et al. NOD2 (CARD15) mutations in Crohn's disease are associated with diminished mucosal alpha-defensin expression. Gut 2004; 53(11):1658-64.

32. Biswas A, Liu YJ, Hao L, Mizoguchi A, Salzman NH, Bevins CL, et al. Induction and rescue of Nod2-dependent Th1-driven granulomatous inflammation of the ileum. Proc Natl Acad Sci USA 2010; 107(33):14739-44.

33. Strober W, Watanabe T. NOD2, an intracellular innate immune sensor involved in host defense and Crohn's disease. Mucosal Immunol 2011; 4(5):484-95.

34. Abreu MT, Taylor KD, Lin YC, Hang T, Gaiennie J, Landers CJ, et al. Mutations in NOD2 are associated with fibrostenosing disease in patients with Crohn's disease. Gastroenterology 2002; 123:679-88.

35. Cuthbert AP, Fisher SA, Mirza MM, King K, Hampe J, Croucher PJ, et al. The contribution of NOD2 gene mutations to the risk and site of disease in inflammatory bowel disease. Gastroenterology 2002; 122:867-74.

36. Hampe J, Grebe J, Nikolaus S, Solberg C, Croucher PJ, Mascheretti S, et al. Association of NOD2 (CARD 15) genotype with clinical course of Crohn's disease: A cohort study. Lancet 2002; 359:1661-5.

37. Zhou Z, Lin XY, Akolkar PN, Gulwani-Akolkar B, Levine J, Katz S, et al. Variation at NOD2/CARD15 in familial and sporadic cases of Crohn's disease in the Ashkenazi Jewish population. Am J Gastroenterol 2002; 97:3095-101.

38. Cho JH. The genetics and immunopathogenesis of inflammatory bowel disease. Nat Rev Immunol 2008; 8(6):458-66.

39. Klionsky DJ, Emr SD. Autophagy as a regulated pathway of cellular degradation. Science 2000; 290(5497):1717-21.

40. Kirkegaard K, Taylor MP, Jackson WT. Cellular autophagy: surrender, avoidance and subversion by microorganisms. Nat Rev Microbiol 2004; 2(4):301-14.

41. Levine B. Eating oneself and uninvited guests: autophagyrelated pathways in cellular defense. Cell 2005; 120(2):159-62. 42. Schmid D, Pypaert M, Münz C. Antigen-loading compartments for major histocompatibility complex class II molecules continuously receive input from autophagosomes. Immunity. 2007;26(1):79-92.

43. Hampe J, Franke A, Rosenstiel P, Till A, Teuber M, Huse K, et al. A genome-wide association scan of nonsynonymous SNPs identifies a susceptibility variant for Crohn disease in ATG16L1. Nat Genet 2007; 39(2):207-11.

44. Rioux JD, Xavier RJ, Taylor KD, Silverberg MS, Goyette P, Huett A, et al: Genome-wide association study identifies new susceptibility loci for Crohn disease and implicates autophagy in disease pathogenesis. Nat Genet 2007; 39:596-604.

45. Marcuzzi A, Bianco AM, Girardelli M, Tommasini A, Martelossi S, Monasta L, et al. Genetic and functional profiling of Crohn's disease: autophagy mechanism and susceptibility to infectious diseases. Biomed Res Int 2013; 2013:297501.

46. Cooney R, Baker J, Brain O, Danis B, Pichulik T, Allan P, et al. NOD2 stimulation induces autophagy in dendritic cells influencing bacterial handling and antigen presentation. Nat Med 2010; 16(1):90-7.

47. Travassos LH, Carneiro LA, Ramjeet M, Hussey S, Kim YG, Magalhães JG, et al. Nod1 and Nod2 direct autophagy by recruiting ATG16L1 to the plasma membrane at the site of bacterial entry. Nat Immunol 2010; 11:55–62.

48. Kuballa P, Huett A, Rioux JD, Daly MJ, Xavier RJ. Impaired autophagy of an intracellular pathogen induced by a Crohn's disease associated ATG16L1 variant. PLoS One 2008; 3(10):e3391

49. Cadwell K, Liu JY, Brown SL, Miyoshi H, Loh J, Lennerz JK, et al. A key role for autophagy and the autophagy gene Atg1611 in mouse and human intestinal Paneth cells. Nature 2008; 456(7219):259-63

50. Parkes M, Barrett JC, Prescott NJ, Tremelling M, Anderson CA, Fisher SA, et al. Sequence variants in the autophagy gene IRGM and multiple other replicating loci contribute to Crohn's disease susceptibility. Nat Genet 2007; 39:830-2.

51. Taylor GA, Feng CG, Sher A. p47 GTPases: regulators of immunity to intracellular pathogens. Nat Rev Immunol 2004; 4(2):100-9.

52. Singh SB, Davis AS, Taylor GA, Deretic V. Human IRGM induces autophagy to eliminate intracellular mycobacteria. Science 2006; 313(5792):1438-41.